- Meloun B, Moravek L and Kostka V, Complete amino acid sequence of human serum albumin. FEBS Lett 58: 134-137, 1975.
- Sudlow G, Birkett DJ and Wade DN, The characterization of two specific binding sites on human serum albumin. Mol Pharmacol 11: 824-832, 1975.
- 8. Leo A, Hansch C and Elkins D, Partition coefficients and their uses. Chem Rev 71: 525-616, 1971.
- Bird AE and Marshall AC, Correlation of serum binding of penicillins with partition coefficients. Biochem Pharmacol 16: 2275-2290, 1967.
- Chignell CF, Optical studies of drug-protein complexes.
 III. Interaction of flufenamic acid and other N-arylanthranilates with serum albumin. Mol Pharmacol 5: 455-462, 1969.
- 11. Fujita T, Hydrophobic bonding of sulphonamide drugs with serum albumin. *J Med Chem* 15: 1049–1056, 1972.
- 12. Lucek RW and Coutinho CB, The role of substituents in the hydrophobic binding of the 1,4-benzodiazepines

- by human plasma proteins. Mol Pharmacol 12: 612-619, 1976.
- Seydel JK and Schaper KJ, Quantitative structurepharmacokinetic relationships and drug design. *Phar*macol Ther 15: 131-182, 1982.
- Wanwimolruk S, Birkett DJ and Brooks PM, Structural requirements for drug binding to site II on human serum albumin. Mol Pharmacol 24: 458-463, 1983.
- Zaton A, Martinez A and De Gandarias JM, The binding of thioureylene compounds to human serum albumin. Biochem Pharmacol 37: 3127-3131, 1988.
- Otagiri M, Masuda K, Imai T, Imamura Y and Yamasaki M, Binding of pirprofen to human serum albumin studied by dialysis and spectroscopy techniques. *Biochem Pharmacol* 38: 1-7, 1989.
- Bos OJM, Fischer MJE, Wilting J and Janssen LHM, Mechanism by which warfarin binds to human serum albumin. Stopped-flow kinetic experiments with two large fragments of albumin. *Biochem Pharmacol* 38: 1979-1984, 1989.

Biochemical Pharmacology, Vol. 41, No. 1, pp. 151-153, 1991. Printed in Great Britain.

0006-2952/91 \$3.00 + 0.00 © 1990. Pergamon Press plc

Inhibition of serine esterases in different rat tissues following inhalation of soman

(Received 7 December 1989; accepted 29 August 1990)

The acute toxic effects of organophosphates (OPs) are due to inhibition of acetylcholinesterase (AChE) (EC 3.1.1.7). The inhibition of other serine esterases, such as butyrylcholinesterase (BuChE) (EC 3.1.1.8) and carboxylesterase (CarbE) (EC 3.1.1.1) does not induce any known physiological alterations. Recently, BuChE has been shown to coregulate acetylcholine lifetime in canine trachealis muscle and may therefore in some tissues play a role in the breakdown of acetylcholine (ACh) [1]. BuChE and especially CarbE may, however, be important for detoxification of low doses of OPs in rodents, since they have a high plasma concentration of CarbE [2-4].

A good correlation between the concentration of plasma CarbE and the LD₅₀ of the OP soman in the developing rat has previously been shown [5]. Furthermore, injection of partially purified rat liver CarbE into 14-day-old rats increased the tolerance to soman, indicating that CarbE in plasma may be of great importance for the detoxification of organophosphorus compounds. CarbE may thereby function as a very important barrier which limits the distribution of the toxic agent to vital organs [6], since the difference in plasma concentration of CarbE between different species correlates with the difference in LD₅₀ [7].

The aim of the present work was to elucidate whether CarbE in respiratory tissue and plasma plays an important role in detoxification of soman during inhalation exposure.

Materials and Methods

Chemicals. [1-14C]Acetylcholine chloride ([14C]ACh) was purchased from Amersham International (Bucks, U.K.). Ethopropazine (10-[2-(diethylamino) propyl]phenothiazine) and 4-nitrophenyl butyrate were from the Sigma Chemical Co. (Poole, U.K.). Soman (O-[1,2,2-trimethyl-propyl]-methyl-phosphonofluoridate), assessed to be more than 99% pure by nuclear magnetic resonance

spectroscopy, was synthesized in our laboratory. All other chemicals were of analytical laboratory reagent grade.

Inhalation method. Whole body exposures of male Wistar rats (200–300 g) (Møllegard, Copenhagen) to sub-acute concentrations of the acetylcholinesterase inhibitor soman were carried out in a dynamic inhalation system designed specifically for the exposure of small rodents to highly toxic gases [8]. Two rats were exposed simultaneously in a glass chamber of 2200 mL, the atmospheric concentration of soman was measured by gas chromatography (Carlo Erba, HRGC 5160) with a nitrogen/phosphorus detector.

No symptoms of poisoning were observed during the inhalation period.

Enzyme activity assays. The total cholinesterase (ChE) activity was determined by the radiochemical method of Sterri and Fonnum [9] at 30°. The AChE activity was measured after inhibition of BuChE by 0.2 mM ethopropazine [10]. The CarbE activity was measured by the spectrophotometric method of Ljungquist and Augustinsson [11] with modifications [12]. The protein concentration was determined by the method of Lowry et al. [13].

Means and standard error of the mean (SEM) were calculated for all data. The Student's t-test was used to assess the significance of the differences between data groups.

Results and Discussion

The airways and lungs are the first tissues exposed to toxic gases and vapours and are also the primary uptake sites for some OPs. The results from this study show that long-term exposure to low concentrations of soman primarily inhibits the cholinesterases of the respiratory tissue, plasma and erythrocytes and the CarbE of plasma and airways (Table 1). Although there were no symptoms

Table 1. Activity of AChE, BuChE and CarbE in different tissues in the rat following 40 hr inhalation exposure to two different concentrations of the organophosphorus anticholinesterase soman

Antonia de de la companya de la comp	Ct (soman) ± SEM	ere den ere er	en e	Mean per cent enzyme activity ± SEM	me activity ± SEM	Material debate and the second se	
Enzyme	(mg min/m³)	Airways	Lung	Diaphragm	Brain	Plasma	Erythrocyte
	0	100 ± 9	100 ± 8	100 ± 13	100 + 11	era de la companya d	100 ± 10
Ş	1 4 7	(8.82) N = 14	(3.17) N = 14	(3.83) N = 14	(49.2) N = 7	**************************************	(1.95) N = 12
AChE	128 ± 25	15±3	17±3	95 ± 14	123 ± 17		15 ± 3
	,	21 = N = 12	*** N = 12	NS N = 12	8 = X SX	nasjadan,	** N = 12
	560 ± 77	2+1	5±2	26 ± 7	59 ± 12		8 + 3
		6 = N ***	6=N**	6 = N ***	* N = 7)-was	V= N ***
	0	100 ± 11	100 ± 8	100 ± 17	100 ± 22	100 ± 15	
		(17.5) N = 14	(4,78) N = 14	(1.55) N = 14	(8.45) N = 7	(1.34) N = 14	Addresses
BuChE	128 ± 25	7+3	37±5	31 ± 12	50 ± 18	31 ± 9	
		*** N = 12	*** N = 12	** N = 12	NS N = 8	** N = 12	apanopani.
	560 ± 77	0	8+3	41 ± 11	65 ± 23	8+3	
		0 ∥ Z ***	6 N ***	6 Z *	NS N = 7	6 Z ***	ana application
	•	100 ± 15	100 ± 10	100 ± 22	100 ± 9	100 ± 8	
		(280) N = 14	(431) N = 14	(102) N = 14	(60.2) N = 7	(81.8) N = 14	-
CarbE	128 ± 25	53 ± 12	63 ± 5	81 ± 12	97,≠9	39 ± 6	
		* N = 12	** N = 12	NS N = 12	NS N = 8	** N 12	*******
	560 ± 77	25 ± 5	8 + 69	59 ± 12	102 ± 7	12 ± 2	
		6 " Z **	6 ≈ X *	6 = N SN	NSN = 7	6 11 × ***	Addispers

Activities of acetylcholinesterase (AChE), butyrylcholinesterase (BuChE) and carboxylesterase (CarbE) in per cent of control (unexposed rats) after exposure to 0.05 and 0.2 mg/m³ (40 hr) of soman. Specific activity (nmol/min/mg protein) is given in parentheses for each control. The enzyme activities were determined as described in Materials and Methods. Values represent mean \pm SEM of N animals.

*** P < 0.001; ** P < 0.05; NS, $P \ge 0.05$.

of poisoning observed during the inhalation experiments, also the cholinesterases in the diaphragm were inhibited to a large extent by soman.

The inhalation experiments (40 hr) with two concentrations of soman (128 \pm 12 and 560 \pm 77 mg min/m³) inhibited the AChE activity in the airways (85 and 98%, respectively), lung (83 and 95%, respectively) and erythrocytes (85 and 92%, respectively) to approximately the same extent. The BuChE activity in lung (63 and 92%, respectively) and plasma (69 and 92%, respectively) were inhibited to approximately the same extent after the soman exposure, although BuChE was apparently less inhibited than AChE. Inhibition of AChE and BuChE were less pronounced in the diaphragm and the brain at both concentrations of soman than in the other tissues examined. This difference in the degree of inhibition is probably due to detoxification by covalent binding of soman to the active site of CarbE and BuChE before soman actually reaches these target tissues [2, 3, 4, 14, 15]. Furthermore, hydrolysis of soman in the liver by phosphorylphosphatases limits its accumulation in the blood [16].

The CarbE activities in diaphragm and brain were not significantly inhibited by soman. The CarbE activity in plasma, however, was significantly (P < 0.001) inhibited (61 and 88%, respectively) following exposure to both the high and the low concentration of soman as were the CarbE activities in airways (47 and 75%, respectively) and lung (37 and 31%, respectively). There are two main groups of CarbE in plasma that can be separated on the basis of their specificity to methyl butyrate and 4-nitrophenyl butyrate [12]. It has been suggested that the plasma CarbE with the highest specificity for 4-nitrophenyl butyrate is the most important enzyme for detoxification after injection of soman [5, 6, 12].

Our results show that inhaled soman inhibits plasma CarbE significantly more than the CarbE in lung and airways. Separate studies have shown that there is only a small difference in the bimolecular inhibition constants of plasma and lung CarbE with soman (R. Gaustad, NDRE, personal communication). Since the cholinesterases of the respiratory tissues, including lung, and the blood were equally inhibited, the small reduction of the CarbE activity in lung compared to plasma was surprising. One possible explanation is the cellular localization of the enzymes. It may be that lung CarbE is not readily available to soman. A similar difference in the level of inhibition between ChE and CarbE is seen in rat liver after in vitro perfusion with soman [17].

In summary, the potent inhibition of CarbE activity in plasma during long-term (40 hr) sub-acute inhalation exposure to soman indicates that CarbE in plasma represents a very important barrier to sub-acute concentrations of inhaled soman. The CarbEs in the airways and the lung are inhibited to a lower extent and thus seems not to be as important as plasma CarbE. The CarbEs in the respiratory tissue are, however, more inhibited after inhalation exposure compared to after injection of soman [12], and may accordingly play a more important role in the detoxification of inhaled soman.

Acknowledgements-The authors are grateful to Ms Rita Tansø for her excellent technical assistance and to Mr John Tørnes for the determination of soman concentrations in the inhalation system. This work was supported by the US Army Medical Research and Development Command (Contract nr. DAMD 17-87-G-7004).

Norwegian Defence Research Establishment Division for Environmental **Toxicology** 2007 Kieller Norway

PER WALDAY* Pål Aas FRODE FONNUM

REFERENCES

- 1. Adler M, Moore D and Filbert MG, Regulation of acetylcholine hydrolysis in airway smooth muscle. Proceedings from NATO RSG-3, Panel VIII, Meeting in Washington DC, U.S.A. pp. 457-473, 1988.
- 2. Fonnum F and Sterri SH, Factors modifying the toxicity of organophosphorus compounds including soman and sarin. Fund Appl Toxicol 1: 143-147, 1981.
- 3. Sterri SH, Lyngaas S and Fonnum F, Toxicity of soman after repetitive injection of sublethal doses in guineapig and mouse. Acta Pharmacol Toxicol 49: 8-13, 1981.
- 4. Sterri SH, Factors modifying the toxicity of organophosphorus compounds including dichlorvos. Acta Pharmacol Toxicol 49 (Suppl V): 67-71, 1981.
- 5. Sterri SH, Berge G and Fonnum F, Esterase activities and soman toxicity in developing rat. Acta Pharmacol Toxicol 57: 136-140, 1985.
- 6. Fonnum F, Sterri SH, Aas P and Johnsen H, Carboxylesterases, importance for detoxification of anticholinesterases organophosphorus and chothecenes. Fund Appl Toxicol 5: S29-S38, 1985.
- 7. Sterri SH and Fonnum F, Carboxylesterase—the soman scavenger in rodents; hererogeneity and hormonal influence. In: Enzymes Hydrolysing Organophosphorus Compounds (Eds. Reiner E, Aldridge WN and Hoskin FCG), pp. 155-164. Ellis Horwood, Chichester, 1989.
- 8. Aas P, Sterri SH, Hjermstad HP and Fonnum F, A method for generating toxic vapors of soman: toxicity of soman by inhalation in rats. Toxicol Appl Pharmacol 80: 437-445, 1985.
- 9. Sterri SH and Fonnum F, Isolation of organic anions by extraction with liquid anion exchangers and its application to micromethods for acetylcholinesterase and 4-aminobutyrate aminotransferase. Eur J Biochem 91: 215-222, 1978.
- 10. Todrik A, The inhibition of cholinesterases by antagonists of acetylcholine and histamine. Br J Pharmacol 9: 76-83, 1954.
- 11. Ljungquist Å and Augustinsson K-B, Purification and properties of two carboxylesterases from rat liver microsomes. Eur J Biochem 23: 303-313, 1971.
- 12. Sterri SH, Johnsen BA and Fonnum F, A radiochemical assay method for carboxylesterase, and comparison of enzyme activity towards the substrates methyl[1-¹⁴C]butyrate and 4-nitrophenyl butyrate. Biochem Pharmacol 34: 2779-2785, 1985.
- 13. Lowry OH, Rosebrough NJ, Farr AL and Randall RJ, Protein measurement with the Folin phenol reagent. J Biol Chem 193: 265-275, 1951.
- 14. Jansz HS, Posthumus CH and Cohen JA, On the active site of horseliver ali esterase I. Reaction of the enzyme with diisopropyl phosphorofluoridate. Biochim Biophys Acta 33: 387-395, 1959.
- 15. Jansz HS, Posthumus CH and Cohen JA, On the active site of horseliver ali esterase II. Amino acid sequence in the DFP-binding site of the enzyme. Biochim Biophys Acta 33: 396-403, 1959.
- 16. Little JS, Broomfield CA, Fox-Talbot MK, Boucher LJ, MacIver B and Lenz DE, Partial characterization of an enzyme that hydrolyzes sarin, soman, tabun, and diisopropyl phosphorofluoridate (DFP). Biochem Pharmacol 38: 23-29, 1989.
- 17. Sterri SH, Valdal G, Lyngaas S, Odden E, Malthe-Sørenssen D and Fonnum F, The mechanism of soman detoxification in perfused rat liver. Biochem Pharmacol 32: 1941-1943, 1983.

^{*} Correspondence to: P. Walday, Norwegian Defence Research Establishment, Division for Environmental Toxicology, PO Box 25, N-2007 Kjeller, Norway.